Inhibition of Histamine-Induced Human Conjunctival Epithelial Cell Responses by Ocular Allergy Drugs

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**Objective:** To evaluate the effects of topical ocular drugs with histamine H1-antagonist activity on histamine-stimulated phosphatidylinositol turnover and interleukin (IL) 6 and IL-8 secretion from human conjunctival epithelial cells.

**Methods:** Primary human conjunctival epithelial cell cultures were stimulated with histamine in the presence or absence of test drugs. Phosphatidylinositol turnover was quantified by ion exchange chromatography and cytokine content of supernatants by enzyme-linked immunosorbent assay.

**Results:** Antazoline hydrochloride, emedastine difumarate, levocabastine hydrochloride, olopatadine hydrochloride, and pheniramine maleate attenuated histamine-stimulated phosphatidylinositol turnover and IL-6 and IL-8 secretion. Emedastine was the most potent in ligand binding, phosphatidylinositol turnover, and IL-6 secretion, with dissociation constant and 50% inhibitory concentrations of 1-3 nmol/L. Olopatadine, antazoline, and pheniramine exhibited similar H1-binding affinities (32-39 nmol/L). However, olopatadine was approximately 10-fold more potent as an inhibitor of cytokine secretion (50% inhibitory concentration, 1.7-5.5 nmol/L) than predicted from binding data, while antazoline and pheniramine were far less potent (20- to 140-fold) in functional assays. Levocabastine (dissociation constant, 52.6 nmol/L) exhibited greater functional activity (50% inhibitory concentration, 8-25 nmol/L) than either antazoline or pheniramine.

**Conclusions:** Histamine-stimulated phosphatidylinositol turnover and cytokine secretion by human conjunctival epithelial cells are attenuated by compounds with H1-antagonist activity. However, antihistaminic potency alone does not predict anti-inflammatory potential. Olopatadine, emedastine, and levocabastine were notably more potent than pheniramine and antazoline.

**Clinical Relevance:** Selected topical ocular drugs with antihistaminic activity may offer therapeutic advantages to patients with allergic conjunctivitis by inhibiting proinflammatory cytokine secretion from human conjunctival epithelial cells.


Human conjunctival epithelial cells (HCEs) secrete cytokines after stimulation by various cell-activating agents. Gamache et al showed that primary cultures of HCEs secrete tumor necrosis factor α, interleukin (IL) 6, IL-8, and granulocyte-macrophage colony-stimulating factor (GM-CSF) after IL-1α, phorbol myristate acetate, and calcium ionophore A23187 treatment. The authors suggested that this capability of the conjunctival epithelium indicates a possible effector function for the tissue in allergic conjunctivitis. Other investigators have reported that histamine, a major mediator of allergic diseases, induces the production of cytokines by airway epithelium. For instance, GM-CSF release from human tracheal epithelial cells was significantly enhanced by exposure to 1-µmol/L histamine. In addition, histamine caused a dose-dependent stimulation of IL-6, IL-8, and GM-CSF release by normal and transformed human bronchial epithelial cells that appeared to occur via histamine H1-receptor activation. The proinflammatory properties of these cytokines are well documented. Elevated IL-6 levels have been reported to be associated with a variety of inflammatory conditions, including asthma, psoriasis, uveitis, and allergic rhinitis. Intravitreal injection of IL-6 has been shown to produce uveitis in rats and rabbits. Interleukin 8 is a potent member of the C-X-C family of chemokines. It promotes integrin expression, neutrophil degranulation, and chemotaxis of basophils and eosinophils.

The presence of these polymorphonuclear leukocytes (primarily eosino-
MATERIALS AND METHODS

CELL CULTURES

Methods detailing the preparation of primary epithelial cell cultures and cytokine release studies with the use of these cells have been described. Briefly, cultures of HCEs were initiated from donor tissues obtained by various eye banks within 8 hours post mortem. The tissues were enzymatically digested overnight. Epithelial cells were gently scraped from the tissue surface, dissociated into a single cell suspension, and cultured in Clonetics keratinocyte growth medium (Biowhittaker Corp, Walkersville, Md). Cells were used only through passage 6. Cultures were maintained in a preconfluent state to prevent differentiation. Cells were identified as epithelial by positive keratin staining, as described previously.

CYTOKINE ASSAYS

Several compounds with histamine H1-antagonist activity were evaluated for their ability to inhibit secretion of cytokines (IL-6 and IL-8) from cultured HCEs in response to histamine stimulation. Cells were plated at 2 x 10⁴ cells per well and cultured overnight in 5% carbon dioxide at 37°C. The following day, keratinocyte growth medium containing test compound was added directly to wells and the cells were incubated for 30 minutes before 24-hour stimulation with histamine (30 μmol/L). Three culture wells were used for each treatment group. At harvest, cell monolayers were examined microscopically to confirm viability and supernatants were collected, centrifuged at 200g, and stored at -20°C. Samples were analyzed for IL-6 and IL-8 by enzyme-linked immunosorbent assay (R&D Systems, Minneapolis, Minn) as directed by the manufacturer. The sensitivities of each enzyme-linked immunosorbent assay are 0.7 pg/mL for IL-6 and 3.0 pg/mL for IL-8.

HISTAMINE-INDUCED PHOSPHATIDYLINOSITOL TURNOVER

The determination of phosphatidylinositol (PI) turnover induced by stimulation of phospholipase C in HCEs was performed as previously described with minor modifications. The HCEs were incubated with tritiated ([¹³H]) myoinositol (0.037 MBq/0.5 mL; 55.3-62.9 x 10⁶ Bq/mmol; Amersham Life Science, Inc, Arlington Heights, Ill) in Dulbecco modified Eagle medium (GIBCO, Gaithersburg, Md) for 24 hours in 5% carbon dioxide at 37°C to label the cell membrane lipids. Cells were then exposed to histamine (10 nmol/L to 1 mmol/L) for 60 minutes at 23°C. To determine the potencies of the antagonists, the drugs were added to the cells 20 minutes before the addition of histamine (100 μmol/L). The assay was terminated by the addition of ice-cold 0.1-mol/L formic acid. With ion exchange columns containing 1 mL of AG1-X8 resin in formate form, free [¹³H]myoinositol was removed from the cell lysates with deionized water; the water-soluble [¹³H]inositol phosphates were then eluted with 1.2-mol/L ammonium formate. The [¹³H]inositol phosphates were quantified by liquid scintillation spectrometry.

DATA ANALYSIS

The antagonist potency (IC₅₀) was defined as the concentration of the drug required to produce 50% inhibition of the agonist-stimulated functional response. Data derived from the cytokine assays were calculated as mean and SEM values that represent the variability among identically treated culture wells. The dose-dependent effect of pharmacological agents and IC₅₀s were determined by linear regression. Data obtained in the PI turnover assays were analyzed by means of a nonlinear, iterative curve fitting program as previously described. Data are expressed as mean ± SEM from 3 to 5 independent experiments.

TEST COMPOUNDS

Compounds were obtained as follows: antazoline hydrochloride and pheniramine maleate (Sigma-Aldrich Corp, St Louis, Mo); emedastine difumarate (Kanebo Ltd, Osaka, Japan); olopatadine hydrochloride (Kyowa Hakko Kogyo Co Ltd, Tokyo, Japan); and levocabastine hydrochloride (Livostin; Ciba Vision Ophthalmics, Atlanta, Ga). Histamine dihydrochloride was obtained from Research Biochemicals International, Natick, Mass.

HCEs. These receptors were coupled to inositol phosphate generation, which mobilized intracellular calcium. Calcium mobilization peaked within 10 seconds and was sustained for 20 minutes after stimulation with histamine. Pharmacological studies indicated that histamine H₁-receptor antagonists potently antagonized these effects. Recently, Weimer et al demonstrated that histamine stimulation of HCEs induced the secretion of IL-6, IL-8, and GM-CSF in a concentration- and time-dependent manner. They also presented evidence that the potent H₁-antagonist emedastine inhibited cytokine secretion at concentrations consistent with its published affinity for the H₁ receptor.

The current experiments were conducted to compare the inhibitory effects of compounds with H₁-antagonist activity currently available for topical ocular use on histamine-stimulated activation of the second messenger system and cytokine release from HCEs.
RESULTS

Exposure of HCEs to 100-µmol/L histamine maximally stimulated PI turnover (2.54 ± 0.16-fold above basal levels). Similarly, exposure of these cells to 30-µmol/L histamine increased IL-6 and IL-8 secretion 1.59 ± 0.19- and 1.80 ± 0.28-fold above basal levels, respectively. (Basal levels of the cytokines were 7667 ± 2110 pg/10^6 cells [n = 4] for IL-6 and 9857 ± 2386 pg/10^6 cells [n = 6] for IL-8.)

Treatment of HCEs with drugs possessing antihistaminic activity and available for topical ocular administration before histamine exposure resulted in concentration-dependent inhibition of PI turnover, IL-6 secretion, and IL-8 secretion. All 5 compounds tested produced concentration-dependent inhibition of the histamine-stimulated cell functional responses (Figure 1, Figure 2, and Figure 3).

Emedastine was the most potent compound tested (Table). The potency of emedastine in intact cells was consistent with its activity determined in receptor binding assays with the use of tissue homogenates. Its IC_{50} values for histamine-induced PI turnover and IL-6 and IL-8 secretion were 1.54, 2.5, and 4.0 nmol/L, respectively. Levocabastine and olopatadine were also potent inhibitors of these histamine-stimulated responses. Levocabastine inhibited the PI turnover and IL-6 and IL-8 secretion (IC_{50}s, 8.32, 25.1, and 11.9 nmol/L, respectively). Olopatadine was more potent than predicted from its published histamine H_{1}-receptor binding affinity (36 nmol/L),\(^2\) with IC_{50}s of 10.03, 5.5, and 1.7 nmol/L for PI turnover and IL-6 and IL-8 secretion, respectively. In fact, olopatadine inhibited histamine-stimulated secretion of IL-8 at a concentration (IC_{50}, 1.7 nmol/L) lower than emedastine’s efficacious concentration in the same assay system.

Antazoline and pheniramine, 2 first-generation topical ocular antihistamines, were dramatically less potent inhibitors of these histamine-induced cell-based responses (PI turnover, IL-6 and IL-8 secretion) than predicted from their histamine H_{1}-receptor binding affinities (Table). The calculated IC_{50} values for these compounds on the parameters listed above ranged from

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**Figure 1.** Effect of histamine H_{1}-antagonists on histamine-induced phosphoinositide (PI) turnover in human conjunctival epithelial cells. Tritiated ([\(^3\)H]myoinositol–labeled cells were exposed to drug for 20 minutes before stimulation with histamine for 60 minutes. [\(^3\)H]inositol phosphates were quantified by ion exchange chromatography followed by liquid scintillation spectrometry.

**Figure 2.** Effect of histamine H_{1}-antagonists on histamine-induced interleukin 6 secretion from human conjunctival epithelial cells. Cells were incubated with test compound for 30 minutes before stimulation with histamine (30 µmol/L) for 24 hours. Supernatants were analyzed for cytokines by specific enzyme-linked immunosorbent assay.

**Figure 3.** Effect of histamine H_{1}-antagonists on histamine-stimulated interleukin 8 secretion from human conjunctival epithelial cells. Assay and data collection procedures are as noted in Figure 2.
Histamine H<sub>1</sub> Antagonists: Inhibition of IL-6 and IL-8 Secretion and PI Turnover in Human Conjunctival Epithelial Cells and H<sub>1</sub> Receptor Binding Affinities*  

<table>
<thead>
<tr>
<th>H&lt;sub&gt;1&lt;/sub&gt; Antagonist</th>
<th>IL-6 IC&lt;sub&gt;50&lt;/sub&gt;, nmol/L</th>
<th>IL-8 IC&lt;sub&gt;50&lt;/sub&gt;, nmol/L</th>
<th>PI Turnover IC&lt;sub&gt;50&lt;/sub&gt;, nmol/L</th>
<th>Binding K&lt;sub&gt;i&lt;/sub&gt;, nmol/L</th>
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<tr>
<td>Emedastine difumarate</td>
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<td>4.0</td>
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<td>Pheniramine maleate</td>
<td>4826.0</td>
<td>1216.0</td>
<td>4500.00</td>
<td>33.90‡</td>
</tr>
</tbody>
</table>

*IL indicates interleukin; PI, phosphoinositide; IC<sub>50</sub>, 50% inhibitory concentration; and K<sub>i</sub>, the concentration of the drug required at equilibrium to inhibit the receptor binding by 50%.
†From Sharif et al.20
‡From Yanni et al.22

Histamine is recognized as a primary mediator of allergic disease. Its acute vascular effects lead to erythema and edema, and its pruritogenic effects are responsible for the itch characteristic of allergic conjunctivitis.23 Additional biological effects of histamine have been reported. The most interesting of these relative to its role as a mediator of allergic diseases is its ability to stimulate or up-regulate proinflammatory cytokine synthesis and/or secretion. Delneste et al24 investigated the effect of histamine on adhesion molecule expression and IL-6 production by human vascular endothelial cells. The authors reported that histamine at concentrations ranging from 10 µmol/L to 1 nmol/L increased IL-6 synthesis from these cells. These authors also reported that IL-8 messenger RNA expression and secretion were enhanced by exposure of endothelial cells to histamine in concentrations greater than 1 µmol/L.25 Similar findings by Tonnel et al26 with the use of human umbilical vein endothelial cells indicated that histamine H<sub>1</sub> and H<sub>2</sub> receptors play a role in cytokine secretion.

Histamine has also been reported to stimulate cytokine secretion from epithelial cells. Secretion of IL-6, IL-8, and GM-CSF by bronchial epithelial cells has been demonstrated.3 Noah et al27 suggested that a correlation exists between calcium influx and IL-6 secretion in a bronchial epithelial cell line in response to stimulation with histamine (100 µmol/L). However, using the antihistamines loratadine and cetirizine hydrochloride, Ansellenn et al28 failed to demonstrate an inhibitory effect on cytokine secretion from bronchial epithelial cells. These investigators therefore suggested that histamine does not play a role in cytokine production. Further experimentation has shown that human tracheal epithelial cells do produce GM-CSF after exposure to histamine.3 Recently, Weimer et al29 demonstrated that histamine induces a concentration- and time-dependent secretion of IL-6, IL-8, and GM-CSF from HCEs. These authors presented evidence that the histamine-stimulated cytokine response was the result of histamine H<sub>1</sub>-receptor activation in these cells. These findings are supported by data showing that the effects of histamine on PI turnover and intracellular Ca<sup>2+</sup> concentration in HCEs were not significantly blocked by H<sub>2</sub> and H<sub>3</sub> antagonists but were dramatically reduced by H<sub>1</sub> antagonists.38

Our data confirm histamine’s ability to stimulate PI turnover and cytokine secretion from HCEs. The concentrations of the agonist used in the present studies (30-100 µmol/L) are consistent with previous cytokine-stimulating concentrations of the biogenic amine.2,3,10,12,25

Current data demonstrate that compounds capable of antagonizing histamine H<sub>1</sub> receptors prevent histamine-stimulated IL-6 and IL-8 secretion. These data confirm emedastine’s ability to prevent cytokine secretion39 and characterize the effects of other compounds that possess antihistaminic activity. First-generation topical ocular anti-histamines antazoline and pheniramine have reported affinities for the histamine H<sub>1</sub> receptor of 38.4 and 33.9 nmol/L, respectively.20 The binding paradigms used to generate these values used cell membranes as the receptor source. In the present studies, which used living, whole cells as the test systems, the 2 compounds were surprisingly less potent. This was true not only for cytokine secretion but also for histamine-stimulated PI turnover. These physiological effects are linked via increased intracellular Ca<sup>2+</sup> concentrations, which are known to facilitate secretory events. Supporting data obtained with calcium ionophore A<sub>23187</sub> exposure of HCEs have shown a stimulation of cytokine secretion.1 The IC<sub>50</sub> values for antazoline and pheniramine ranged from 652 to 4200 nmol/L and from 1216 to 4826 nmol/L, respectively. These data suggest that the first-generation anti-histamines have limited ability to interact with intact cells of the human conjunctiva. These findings may explain the limited clinical utility of these early anti-histamines when used as single-entity products without vasoconstrictors.

Second-generation topical ocular anti-histamines, levocabastine and emedastine, inhibited histamine-stimulated PI turnover and cytokine secretion. The potency of these molecules in these assays was consistent with their affinities for the H<sub>1</sub> receptor (52.6 nmol/L for levocabastine and 1.22 nmol/L for emedastine).20 Levocabastine’s IC<sub>50</sub> values ranged from 8.3 to 25.1 nmol/L. Emedastine has been reported to be the most potent anti-histamine available for topical ocular use.20,29 The drug’s histamine H<sub>1</sub>-receptor affinity of 1.22 nmol/L is reflected in the data obtained in the present experiments. Emedastine’s potency in preventing cytokine secretion by HCE may partially explain the advantages noted with this compound during clinical comparative trials with levocabastine.30

Interesting results were obtained with olopatadine. Olopatadine is marketed as Patanol for topical ocular use. The compound is a human conjunctival mast cell degranulation inhibitor and antihistamine.22,31,32 Published reports demonstrate the compound’s antiallergic activity in vivo.31 Results presented herein indicate that the compound is more potent as an inhibitor of cytokine secretion than would have been predicted from its
H1-receptor affinity (36 nmol/L). Olopatadine potentially inhibited secretion of IL-6 and IL-8 from HCEs. The IC50 values for the drug were 5.5 and 1.7 nmol/L, respectively. These IC50 values were also approximately 2-fold and 10-fold lower, respectively, than those predicted from the functional second messenger data. Olopatadine was reported to be more potent as an inhibitor of histamine-enhanced tumor necrosis factor α-stimulated adhesion molecule expression than predicted from the drug's receptor binding affinity. 29 One possible explanation for this increased efficacy is a non-specific antisecretory effect. However, this does not appear to be the case. A report by Ikemura et al 35 presented data showing that olopatadine (1-100 µmol/L) did not inhibit the release of β-glucuronidase from human polymorphonuclear neutrophils after stimulation with the calcium ionophore A23187.

The results presented herein confirm histamine's ability to stimulate PI turnover and cytokine secretion from HCEs. First-generation antihistamines pheniramine and antazoline are dramatically less potent in the whole cell assays used compared with their receptor affinities, possibly providing some insight into their limited clinical utility. The current generation of topical ocular antihistamines, emedastine and levocabastine, exhibit potencies consistent with their receptor affinities (deviation approximately 2-fold). Importantly, olopatadine is 10 times more potent as an inhibitor of histamine-stimulated cytokine secretion than predicted from its histamine H1-receptor affinity. These data suggest that olopatadine may offer additional therapeutic benefits that complement the mast cell stabilization and antihistaminic activities observed in the clinic.

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