Immunohistochemical and Molecular Genetic Evidence for Type IV Collagen α5 Chain Abnormality in the Anterior Lenticus Associated With Alport Syndrome

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Objective: To present evidence for a type IV collagen α5 chain (α5(IV)) abnormality in the anterior lens capsule of a patient with anterior lenticonus associated with Alport syndrome.

Methods: The anterior lens capsule obtained from a 54-year-old man with anterior lenticonus associated with Alport syndrome was examined ultrastructurally and stained immunohistochemically for the α chains of type IV collagen, α1(IV) to α6(IV). A search was also made for a mutation in the COL4A5 complementary DNA encoding the α5(IV) chain by reverse transcription–polymerase chain reaction of illegitimate transcripts.

Results: The anterior lens capsule of the patient was much thinner than that of normal subjects and lacked the α3(IV) to α6(IV) chains immunohistochemically, while control specimens stained positively for all of the α(IV) chains. The patient had a C-to-T transition at nucleotide 5231 causing a nonsense mutation, R1677X, in the COL4A5 complementary DNA.

Conclusion: Our findings demonstrated that normal anterior lens capsules express all of the α(IV) chains and that a patient with anterior lenticonus associated with Alport syndrome had a mutation in the COL4A5 gene resulting in the lack of immunoreactivity to α3(IV) to α6(IV) chains in the anterior lens capsule.

Clinical Relevance: This study showed abnormal composition of α(IV) chains in the anterior lens capsule of a patient with anterior lenticonus caused by a nonsense mutation in the COL4A5 gene. Further investigation of the phenotype-genotype relationship will provide a better understanding of the molecular pathogenesis of anterior lenticonus.

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ELECTRON MICROSCOPY

The central portion of the anterior capsule was used for the ultrastructural study. The specimens were immersed in a phosphate buffer (pH, 7.2-7.4) containing 2.5% glutaraldehyde and postfixed in 1% osmium tetroxide in the same buffer. The specimens were dehydrated in graded ethanol and embedded in epon using standard procedures. Ultrathin sections for electron microscopy were stained with uranyl acetate and lead citrate and were observed by transmission electron microscopy (H-7100; Hitachi, Tokyo, Japan).

IMMUNOHISTOCHEMISTRY

The specimen of the patient with AS was snap-frozen in a dry ice and acetone bath, transferred without thawing, and stored at −70°C for several months until use in the immunohistochemical experiments. The anterior capsules from 2 of the 4 control subjects were processed and stored on the same occasion. The specimens from 2 other control subjects were obtained 2 years later and processed in the same way.

Cryostat sections (3 µm) of the peripheral portion of the anterior capsule were cut at a constant temperature (−25°C) and fixed for 10 minutes in acetone. The sections were placed in an acid buffer made by mixing an equal volume of 0.1M potassium chloride and 0.1M hydrochloric acid (pH, 1.5) for 10 minutes and rinsed 3 times with phosphate-buffered saline (PBS). The sections were then immersed in 10% normal goat serum in PBS for 10 minutes and rinsed 3 times with PBS.

Serial sections were stained by an indirect method using primary antibodies against the α1(IV) to α6(IV) chains prepared as reported previously. Briefly, rat monoclonal antibodies were raised against synthetic peptides of nonconsensus amino acid sequences close to the carboxyl terminus of the noncollagenous (NC) domain of each α(IV) chain by a method using rat medial iliac lymph nodes. The antibodies were diluted 1:10 with PBS containing 1 mg/mL of bovine serum albumin. After a 60-minute incubation with the primary antibodies at room temperature, the sections were rinsed 3 times with PBS. Then the secondary antibody, fluorescein isothiocyanate–conjugated goat F(ab’2) anti-rat IgG (Organon Teknika-Cappel, Durham, NC) was added to the sections. The secondary antibody was diluted 1:50 with PBS containing 5% normal human serum prepared in our laboratory. After a 30-minute incubation, the sections were rinsed 3 times with PBS and mounted on coverglass with 20% glycerin (Wako Pure Chemical Industries, Ltd, Osaka, Japan). The sections were viewed with a reflecting microscope (BH2-RFCA; Olympus Optical Co, Tokyo).

Immunostaining of the specimens from the patient and control subjects was performed simultaneously using the same solutions.

MUTATION ANALYSIS OF THE COL4A5 COMPLEMENTARY DNA

The complementary DNA (cDNA) sequence encoding the noncollagenous (NC1) domain of the α5(IV) chain was determined by reverse transcription–polymerase chain reaction products using illegitimate COL4A5 transcripts as described by Inoue et al. Polyadenylated RNAs were extracted from peripheral leukocytes of each subject and reverse transcribed. The cDNA region encoding the NC1 domain of the α5(IV) chain was amplified by polymerase chain reaction with 2 sets of oligonucleotide-primer pairs. Direct sequencing of the polymerase chain reaction products was performed on an automated DNA sequencer.

RESULTS

ULTRASTRUCTURE

The thickness of the anterior capsule of the patient with AS was 3 µm in the central portion and noticeably thinner than that of the controls (20 µm) (Figure 1). The thickness of a normal anterior capsule has been reported to be 12 to 21 µm. Dehiscences were noted up to two thirds of the depth from the lens epithelium (Figure 1A, arrows) (A, original magnification ×4000; B, original magnification ×2700).

Figure 1. Transmission electron micrographs of the anterior lens capsule from a patient with Alport syndrome associated with anterior lenticonus (A) and of the capsule obtained from an eye with uncomplicated age-related cataract (B). The double-headed arrows indicate the anterior lens capsule. Dehiscences are noted up to two thirds of the depth from the lens epithelia (Figure 1A, arrows) (A, original magnification ×4000; B, original magnification ×2700).
IMMUNOHISTOCHEMICAL STAINING

Figure 2B demonstrates the α1(IV) to α6(IV) chain-like immunoreactivities present throughout the anterior lens capsules in controls. No difference in location of the immunoreactivity between different α chains was observed. In contrast, the lens capsule of the patient with AS was negative for the α3(IV) to α6(IV) chain-like immunoreactivities (Figure 2A). Even undiluted primary antibodies of α3(IV) to α6(IV) chains gave the same results, indicating the absence of immunoreactivity to these chains in the anterior capsule of the patient (data not shown).

The staining patterns of α1(IV) to α6(IV) chains in the lens epithelial cells were identical with the remaining parts of the anterior capsule, although the positive staining might be partly artifactual because of the dissociation of the epithelium during the tissue processing (Figure 2A, arrows).

MUTATION ANALYSIS OF THE COL4A5 cDNA

The patient had a C-to-T transition at nucleotide 5231 resulting in a nonsense mutation, R1677X, in the COL4A5 cDNA (Figure 3). Genomic DNA sequencing of the
The ultrastructural changes of the anterior lens capsule in our patient with AS were in agreement with those described in previous reports. Streeter et al first reported that the anterior lens capsule in AS was markedly thin and showed numerous dehiscences extending up to two thirds of the depth from the lens epithelium. Later, Kato and Junk et al reported similar lesions, with up to two thirds of the depth from the lens epithelium. Takei et al described a female patient whose anterior lens capsule showed normal immunoreactivity to the \( \alpha 5(IV) \) chain-like immunoreactivity and decreased immunoreactivity to the \( \alpha 2(IV) \) chain and decreased immunoreactivity to the \( \alpha 5(IV) \) chain. Therefore, the anterior lens capsule from patients with AS may show abnormal immunostaining of the \( \alpha 3(IV) \) to \( \alpha 5(IV) \) chains, depending on individual genetic defect. However, neither study identified the possible gene mutations responsible for the abnormal \( \alpha (IV) \) chains. In addition, there has been no examination of the \( \alpha 6(IV) \) chain in the anterior lens capsule of patients with AS or of normal subjects.

As demonstrated, the normal anterior lens capsule expressed the \( \alpha 1(IV) \) to \( \alpha 6(IV) \) chains, but only the \( \alpha 1(IV) \) and \( \alpha 2(IV) \) chains were expressed in our patient with AS. Because \( \alpha 1(IV)/\alpha 1(IV)\alpha 2(IV), \alpha 3(IV)/\alpha 4(IV)/\alpha 5(IV), \) and \( \alpha 5(IV)/\alpha 5(IV)/\alpha 6(IV) \) triple helical molecules are the only known components of the type IV collagen network, our results indicate that the normal anterior lens capsule exhibits not only \( \alpha 1(IV)/\alpha 1(IV)/\alpha 2(IV) \) and \( \alpha 3(IV)/\alpha 4(IV)/\alpha 5(IV) \) molecules but also the \( \alpha 5(IV)/\alpha 5(IV)/\alpha 6(IV) \) molecule. Our patient lacked the \( \alpha 3(IV)/\alpha 4(IV)/\alpha 5(IV) \) molecule and the \( \alpha 5(IV)/\alpha 5(IV)/\alpha 6(IV) \) molecule. The most plausible interpretation of our results is that this patient had a defective \( \alpha 5(IV) \) chain.

Recently, several studies have identified a close relationship between the clinical severity of nephritis (phenotype), the immunohistochemical findings of the \( \alpha (IV) \) chains in the glomerular basement membrane, and the genotype of the \( \alpha 5(IV) \) chain. Inoue et al showed that mutations in the NC1 domain of the \( \alpha 5(IV) \) chain were associated with an absence of immunostaining of the \( \alpha 3(IV) \) to \( \alpha 5(IV) \) chains in the glomerular basement membrane and with severe renal dysfunction. Similarly, our patient had renal dysfunction and was negative for the \( \alpha 3(IV) \) to \( \alpha 5(IV) \) (and \( \alpha 6(IV) \)?) chain-like immunoreactivities in the anterior lens capsule. We therefore searched for a mutation in the cDNA sequence corresponding to the NC1 domain of the \( \alpha 5(IV) \) chain and found the nonsense mutation R1677X. This is one of the mutations reported by Inoue at al. The R1677X nonsense mutation is thought to produce a truncated NC1 domain of the \( \alpha 5(IV) \) chain lacking 9 amino acids, including 2 cysteine residues involved in intermolecular cross-linkages, leading to a malformation of collagen network made by the \( \alpha 3(IV)/\alpha 4(IV)/\alpha 5(IV) \) and \( \alpha 5(IV)/\alpha 5(IV)/\alpha 6(IV) \) triple helical molecules. We suggest that the mechanisms causing the defective glomerular basement membrane also caused fragility in the anterior lens capsule in our patient with AS.

Pajari et al reported that the ocular manifestations in AS develop with age but independently of the genotype. However, they identified the gene mutations in only a few of their patients; therefore, the correlation between the genotype and the ocular phenotype in AS needs further investigation. Systematic examinations, in-
cluding electron microscopy, immunohistochemistry, and molecular genetics, in multiple patients with AS with anterior lenticonus will provide a better understanding of the physiological role played by the α(IV) chains in the anterior lens capsule and in the pathogenesis of anterior lenticonus.

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